STRUCTURES OF WITHAMETELIN AND ISOWITHAMETELIN, WITHANOLIDES OF DATURA METEL LEAVES

Subhash C. Sınha, Sujata Kundu, Rakesh Maurya and Anil B. Ray* Department of Medicinal Chemistry, Institute of Medical Sciences Banaras Hindu University, Varanası, India

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Yoshiteru Oshima, Anjana Bagchi and Hiroshi Hikino Pharmaceutical Institute, Tohoku University, Aoba-yama, Sendai, Japan

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ABSTRACT : Structures of withametelin and isowithametelin, two novel hexacyclic withanolides isolated from the leaves of <u>Datura metel</u>, have been established as 1 and 2 respectively, on the basis of detailed spectral and chemical evidences. The facile transformation of withametelin to a ring A aromatic withanolide and some other interesting chemical transformations are described.

The structure elucidation of withametelin, the major steroidal constituent of the dried leaves of <u>Datura metel</u> Linné, was reported by us almost exclusively on the basis of spectral evidence in a preliminary communication.² A full account of the isolation and structure determination of withametelin and its congener, isowithametelin, is presented in this paper together with some interesting chemical transformations of these molecules.

Withametelin, $C_{28}H_{36}O_4$ (M⁺ 436.2426), m.p. 210°, $\lceil \alpha / \rho \rceil$ -64.4°, was proved to bear In its molecule an enone and a α,β -unsaturated- δ -lactone molety from its UV and IR spectral characteristics (λ_{max} 222 nm (ϵ 7000); ν_{max} 1718, 1655 cm⁻¹) which are comparable with those of a typical withanolide.³ Of the five olefinic hydrogen signals discernible in its $^1_{
m H}$ NMR spectrum, the two appearing as singlets with fine splitting at δ 6.76 and 6.02 were assigned to hydrogens of a terminal methylene conjugated with a &-lactone and the remaining three were considered to be associated with steroidal 2,5-dien-1-one system on the basis of their chemical shifts and splitting patterns (δ 6.77, 1H ddd, J=9.8, 5.1, 2.5 Hz, 5.88, 1H dd, J=9.8, 2.5 Hz, 5.58, 1H t, J=5.8 Hz). Chemical evidence to verify the presence of 2,5-dien-1-one molety was secured by acetoxylation of withametelin with mercuric acetate which yielded a derivative ($\underline{3}$) having 6B-acetoxy-2,4-dien-1-one system. This chemical conversion, which can be readily recognized from a change in UV absorption characteristics ($\lambda_{\rm max}$ 310, 207 nm) and appearance of signals for three contiguous olefinic hydrogens (δ 6.94, 1H dd, J=10, 6 Hz, 6.30, 1H dd, J=6, 1 Hz, 6.04, 1H dd, J=10, 1 Hz) and an acetoxy methyl (δ 2.05, 3H s) with concomitant downfield shift of the 19-methyl signal in the 1 H NMR spectrum, has been studied in similar system⁵ and considered to be a diagnostic proof for the presence of the 2,5-dien-1-one molety. The exclusive formation of the 68-acetoxy

derivative ($\underline{3}$) may be rationalized by assuming the formation of a cyclic mercuronium ion⁶ by the addition of ⁺HgOAc on the 5,6-double bond from the less hindered α -face of the molecule followed by the opening of the cyclic complex with concomitant generation of the 4,5-double bond and nucleophilic substitution of the 6α -mercurated compound with acetic acid. A report⁷ of the exclusive formation of 5α , 6α -epoxide of a withasteroid having 2,5-dien-1-one system by treatment with <u>m</u>-chloroperbenzoic acid lent credence to the α -attack, as envisaged in the formation of the cyclic mercuronium complex.

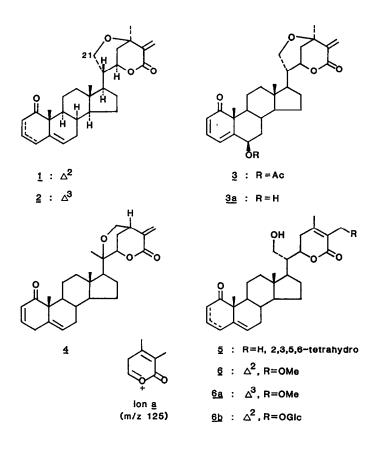
While the isolation of withametelin from a solanaceous plant, its molecular formula and the presence of enone and α,β -unsaturated- δ -lactone chromophores in its molecule, suggested it to be a withanolide or a structurally related compound, it failed to exhibit diagnostic mass and ¹H NMR spectral features of a typical withanolide. The mass spectrum of withametelin showed no significant fragment ion peak formed by fission across C-20,22 bond which is characteristic of common withanolides. Further, the diagnostic double truplet or double doublet splitting pattern of the C-22 carbinyl hydrogen signal which normally appears around δ 4.5, and is regarded as the withanolide "finger print", was not discernible in its 1 H NMR spectrum and instead, a broad 1H singlet was observed at δ 4.65. Again, while a typical withanolide shows, in its ¹H NMR spectrum, five methyl signals including two vinylic methyl signals, only three 3H singlets appeared at δ 0.71, 1.22 and 1.44 in the 1 H NMR spectrum of withametelin and these were assigned respectively, to 18- and 19-methyls and to a methyl on a carbon bearing oxygen function. The absence of two methyl groups in the molecule was, however, compensated by the presence of a terminal methylene (>C=CH $_{n}$) and an oxymethyl (-CH2-O) grouping (δ 3.95, 1H d, J=12.7 Hz, 3.73, 1H dd, J=12.7, 3 Hz). The terminal methylene conjugated to the lactone carbonyl was assigned to C-27 but the location of the oxymethyl group could not be easily settled and it was an open question which of the remaining two methyls (C-21 and C-28) of this $C_{28}^{}$ -steroid was oxygenated?

Of the four oxygen atoms present in withametelin, three are associated with an enone and δ -lactone function and the remaining one was considered to be an ether in view of the fact that withametelin was not amenable to acetylation under normal condition and it exhibited no hydroxyl band in the IR spectrum. Based on these observations and from valency calculation, withametelin was proved to be hexacyclic and the fourth oxygen atom associated with an oxymethyl group was considered responsible for the construction of the sixth ring. The two oxycarbon signals at δ 69.2 (s) and 60.4 (t) in the ¹³C NMR spectrum lent support to this assumption. The foregoing data permitted advancement of two alternative structures 1 and 4 for withametelin. An inspection of these two alternative structures revealed that the cyclic ether is allylic in nature only in 1 and, therefore, it was considered possible to make a correct choice by ascertaining whether the ether oxygen in withametelin is allylic or not. With this objective, withametelin was subjected to catalytic hydrogenation to give a mixture, which on chromatographic resolution yielded a hexahydro derivative (M+ 442) as the major product, along with tetrahydrowithametelin (5) (M⁺ 440) in a poor yield. The tetrahydro derivative (5), formed by hydrogenolysis of the cyclic ether, showed signals for two vinylic methyls (δ 1.93 and 1.88, 3H s each) and the characteristic double triplet for H-22

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(δ 4.37, 1H <u>dt</u>, J=13.5, 3.6 Hz) in its ¹H NMR spectrum and the base peak at <u>m/z</u> 125 (ion <u>a</u>) in its mass spectrum like typical withanolide. Withametelin was thus established to have a cyclic allyl ether function and formulated as <u>1</u> which was verified by ¹³C shift correlation two dimentional spectrum (2D INADEQUATE) and reported in the previous communication.²

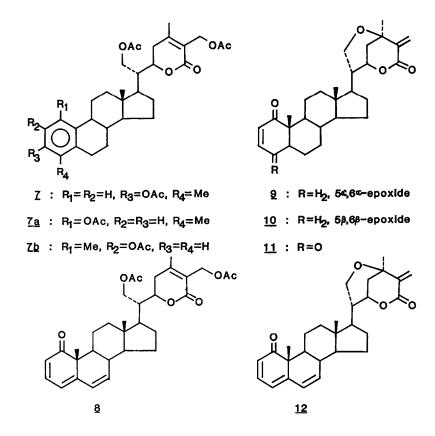
Both tetrahydrowithametelin ($\underline{5}$) and seconthametelin ($\underline{6}$), the latter being obtained in near quantitative yield by treatment of withametelin with methanol in presence of perchloric acid,⁸ showed positive Cotton effect around 250 nm, indicating 22<u>R</u> configuration. A comparative study of the ¹³C NMR spectra of withametelin and its derivatives with that of withaferin A acetate,⁹ revealed 14 α and 178 configurations in withametelin. The stereochemistry at C-20, as shown in the expression <u>1</u> is based on biogenetic arguments because all the reported withanolides unsubstituted at C-20 have the same configuration. The Dreiding stereomodel study of withametelin revealed that 28-methyl and H-22 are on the same side of the molecule.



The smooth addition of methanol to withametelin in presence of perchloric acid⁸ led to the assumption that withametelin, on treatment with acetic anhydride and perchloric acid, should give a diacetate but, to our surprise, the product obtained was a triacetate ($\underline{7}$) (M⁺ 580). An analysis of its spectral properties revealed significant difference in its carbocyclic molety from that of withametelin. The ¹H NMR spectrum of the triacetate ($\underline{7}$) showed signals for three acetoxy methyls ($\delta 2.08$, 2.06 and 2.05, 3H <u>s</u> each), and a vinylic methyl ($\delta 1.98$, 3H <u>s</u>) and the 18-methyl ($\delta 0.76$, 3H <u>s</u>), while the signal for the 19-methyl was found to be missing and it was replaced by a 3H singlet at $\delta 2.30$ assignable to an aromatic methyl group. A pair of 1H doublets at $\delta 7.32$ and 6.80 (J=8 Hz) for two vicinal aromatic hydrogens was discernible in the ¹H NMR spectrum. The presence of two -CH₂-OAc groupings was manifest from two sets of signals appearing as two pairs of doublets ($\delta 4.36$, 1H <u>dd</u>, J=12, 1 Hz, 4.26, 1H <u>dd</u>, J=12, 4 Hz and $\delta 4.88$, 2H AB <u>q</u>, J=15.4 Hz) assigned to 21and 27-methylenes, respectively. The ¹H NMR spectrum of the triacetate ($\underline{7}$) also showed the diagnostic double triplet at $\delta 4.49$ (J=13, 3.5 Hz) for H-22 of withanolides unsubstituted at 20-position.

That the A ring of this triacetate ($\underline{7}$) is aromatic in nature became evident from an inspection of its ¹³C NMR spectrum which lacked the enone carbonyl signal of withametelin and showed six aromatic carbon signals of a tetrasubstituted benzene ring (δ 147.1 <u>s</u>, 138.0 <u>s</u>, 136.8 <u>s</u>, 128.0 <u>s</u>, 123.6 <u>d</u>, 118.9 <u>d</u>). A subtraction UV spectrum of the triacetate ($\underline{7}$) and physalolactone B (1 α -acetoxy-38,20-dihydroxywitha-5,24-dienolide)¹⁰ which showed an absorption maximum at 278 nm also spoke of the presence of a benzenoid chromophore. The mass spectrum of the triacetate ($\underline{7}$) showed not only the molecular ion peak at <u>m/z</u> 580 but also three significant fragment ion peaks at <u>m/z</u> 538, 478 and 418, formed by sequential loss of a molecule of ketene and two molecules of acetic acid. The loss of ketene suggested the presence of a phenol acetate molety which was supported by positive phosphomolybdic acid test.

The aromatization of the ring A of withametelin is assumed to take place by the well known dienone-phenol rearrangement, the dienone in this case being 2,4-dien-1-one, formed by appropriate prototropic shift under the reaction condition. While three possible structures (7, 7a and 7b) having different substitution patterns of the ring A may be conceived, the volume of literature¹¹ available for dienone-phenol rearrangement of related 2,4-dien-3-one steroids suggests the formulation 7 for the triacetate. In fact, that no nOe was detected on aromatic hydrogen signals on irradiation of the aromatic methyl signal at δ 2.30 rules out the substitution pattern as shown in expression 7a. Further, it was found that the 13 C NMR signals of the aromatic carbons of estrone acetate¹² matched well with those of the triacetate, except C-4 and C-6. The signal for the carbon (C-10) corresponding to C-4 of estrone acetate appeared at lower field and that for C-6 at higher field, demonstrating the presence of methyl substituent at C-10 position and thus confirming the correctness of the structure ($\frac{7}{2}$) proposed for the triacetate of withametelin. It is pertinent to mention in this connection that though aromatization of the ring A of withametelin with acetic anhydride and perchloric acid is considered to take place by dienone-phenol rearrangement, withametelin-derived 68acetoxy-2,4-dien-1-one (3) failed to undergo aromatization with acetic acid and perchloricacid and yielded a cojugated trien ($\underline{8}$). While it may be argued that the facile acidcatalyzed elimination of the allylic acetoxy function at C-6 prevents aromatization of the dienone ($\underline{3}$), no suitable explanation is available for the failure in aromatization of the ring A of withametelin on treatment with methanol and perchloric acid. Secowithametelin ($\underline{6}$) obtained under this reaction condition retains the 2,5-dien-1-one system. It is, therefore, quite logical to believe that acetic anhydride has an important role in the aromatization of the ring A of 2,5-dien-1-one steroids.



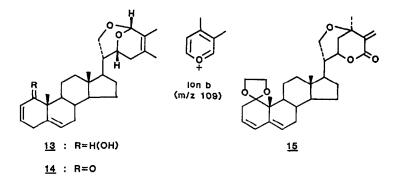
As the variety of oxygen substituents witnessed in AB rings of withanolides are known¹³ to be derived from the one having 2,5-dien-1-one system, efforts were made to introduce some of these oxygen functions in view of their possible existence as natural products. Withametelin on treatment with <u>m</u>-chloroperbenzoic acid yielded a mixture of epoxides. The mixture was resolved by HPLC and the major product, $C_{28}H_{36}O_5$ (M⁺ 452), which showed a 1H narrow triplet at δ 3.02 was recognized as withametelin-5 α ,6 α -epoxide (<u>9</u>) from the negative Cotton effect at 345 nm. The minor product, isomeric with 9 showed positive

Cotton effect at 345 nm and the carbinyl hydrogen signal of the oxirane at $\delta 3.10^{14}$ in its ¹H NMR spectrum and on this basis it was identified as withametelin-58,68-epoxide (<u>10</u>). In an effort to insert a hydroxyl group at the bis-allylic 4-position of withametelin, it was oxidized with selenium dioxide in aqueous acetic acid medium. The reaction mixture, on chromatographic separation yielded four compounds which were identified from detailed spectral analysis as <u>3</u>, <u>3a</u>, <u>11</u> and <u>12</u>.

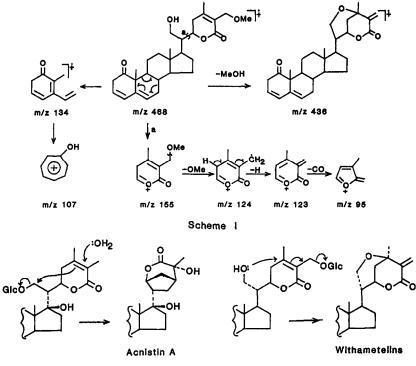
Withametelin, which bears a α -methylene- δ -lactone as well as an oxide ring in its side chain, could be transformed to yet another interesting hexacyclic withanolide (<u>14</u>). Reduction of withametelin with diisobutylaluminium hydride gave an allyl alcohol (<u>13</u>), the formation of which can be explained by hydrogenolysis of allyl ether, reduction of the δ -lactone to lactol and formation of an acetal with the participation of hydroxyl group at C-21. Oxidation of <u>13</u> with pyridinium dichromate in methylene chloride yielded <u>14</u> without affecting the bicyclic system. Mass spectra of both <u>13</u> and <u>14</u> showed a base peak at <u>m/z</u> 109 (ion <u>b</u>) and their ¹H NMR showed the absence of the signals for exocyclic methylene and the low field methyl attached to the carbon bearing oxygen function and instead showed two 3H singlets for two vinylic methyles and a low field 1H broad singlet for the acetal hydrogen.

Daturilin, isolated from the fresh leaves of <u>D</u>. <u>metel</u> by the Pakistani workers¹⁵ has been assigned the same gross structure as withametelin but with 22<u>S</u> configuration. Though no direct comparison of withametelin with daturilin has been made, they appear to be identical and, therefore, 22<u>S</u> configuration assigned to this compound exclusively on the basis of nOe measurements, is not tenable.

Isowithametelin, $C_{28}H_{36}O_4$, m.p. 280°, $\lceil \alpha \rceil_D$ -77.7°, a congener of withametelin, showed a R_f value very close to that of the latter on TLC in a variety of solvent systems and it was found to be present in mother liquor of withametelin from which it was isolated by preparative HPLC work. The UV spectrum of isowithametelin showed twin absorption maxima at 233 and 227 nm, indicating the probable presence of a α,β -unsaturated- δ -lactone and a heteroannular dien chromophore¹⁴ and its IR spectrum showed a strong carbonyl absorption band at 1714 cm⁻¹ but no band for the enone carbonyl or hydroxyl group. The 13 C NMR spectrum of isowithametelin, however, showed two carbonyl carbon signals at δ 210.7 and 165.4 assignable, respectively, to a saturated keto carbonyl and a lactone carbonyl. The ¹H NMR spectrum of isowithametelin showed striking resemblance with that of withametelin, the only observable difference being the chemical shifts and splitting patterns of three olefinic hydrogens of the AB rings and 19-methyl of the two molecules. Based on this finding and from the correspondance of ¹H NMR parameters of these three olefinic hydrogens with those of withanolide I, 16 the AB ring of isowithametelin was proved to have 3,5-dien-1-one system and the structure 2 was advanced for isowithametelin. ¹³C NMR data of isowithametelin (Table I) is in perfect agreement with the proposed structure. Chemical evidence in support of the structure was provided by conversion of withametelin to isowithametelin. When a solution of withametelin in benzene was refluxed with ethylene glycol in presence of p-toluenesulphonic acid, the ketal (15) was obtained as the major product, which on deketalization with \underline{p} -toluenesulphonic acid in aqueous acetone furnished a compound indistinguishable from



isowithametelin in all respects. The methanol adduct ($\underline{6a}$) of isowithametelin obtained by treatment of its methanolic solution with perchloric acid showed positive Cotton effect at 241 nm, indicating that isowithametelin also has 22<u>R</u> configuration like withametelin. The mass spectrum of the methanol adduct ($\underline{6a}$) displayed significant fragment ion peaks, the genesis of which can be very well rationalized from the expression $\underline{6a}$ (Scheme I).



Scheme II

Table I : Carbon-13 shieldings in withametelin and its derivatives (125 MHz, CDCl₂)

Carbo No.	ⁿ <u>1</u>	<u>2</u> *	<u>5</u>	<u>7</u> **	<u>9</u> *	<u>10</u> *	Carbon No.	<u>י</u> 1	<u>2</u> *	5	2	<u>9</u> *	<u>10</u> *
C-1	204.5 <u>s</u>	210.7 <u>s</u>	214.7 <u>s</u>	147.1 <u>s</u>	201.6 <u>s</u>	202.3 <u>s</u>	C-16	26.5 <u>t</u>	26.7 <u>t</u>	26.3 t	27.6 <u>t</u>	25.3 <u>t</u>	25.4 <u>t</u>
C-2	128.0 <u>d</u>	39.6 <u>t</u>	38.4 <u>t</u>	118.9 <u>d</u>	127.6 <u>d</u>	128.2 <u>d</u>	C-17	47.6 <u>a</u>	47.7 <u>a</u>	46.6 <u>d</u>	468 <u>d</u>	46.2 <u>d</u>	46.5 d
C-3	145.0 <u>d</u>	127.0 쇼	27.4 <u>t</u>	123.6 <u>d</u>	141.1 <u>a</u>	143.1 <u>d</u>	C-18	12.7 <u>g</u>	12.9 <u>q</u>	12.6 g	12.2 g	11.7 <u>g</u>	11.5 g
C4	33.4 <u>t</u>	129.5 <u>d</u>	31.2 <u>t</u>	138.0 <u>s</u>	33.0 <u>t</u>	31.9 <u>t</u>	C-19	18.9 g	20.5 g	19.4 g	12.1 g	14.5 g	14.0 g
C-5	135.9 <u>s</u>	141.2 <u>s</u>	141.4 <u>s</u>	136.8 <u>s</u>	63.7 <u>s</u>	61.0 <u>s</u>	C-20	39.9 <u>d</u>	40.0 <u>d</u>	42.6 d	42.7 a	38.8 <u>a</u>	38.6 d
C-6	124.5 <u>d</u>	121.7 <u>đ</u>	121.8 <u>d</u>	27.8 <u>t</u>	57.5 <u>d</u>	62.2 <u>d</u>	C-21	60.4 <u>t</u>	60.6 <u>t</u>	60.0 <u>t</u>	61.8 ±	59.4 t	59.4 t
C-7	30.7 <u>t</u>	31.1 <u>t</u>	32.2 <u>t</u>	27.7 <u>t</u>	27.7 <u>t</u>	30.2 <u>t</u>	C-22	75.8 <u>d</u>	75.7 <u>a</u>	78.2 4	***	74.4 <u>d</u>	74.7 d
C-8	33.2 <u>d</u>	31.8 <u>d</u>	32.0 <u>d</u>	37.7 <u>d</u>	29.7 <u>d</u>	28.8 <u>d</u>	C-23	33.2 <u>t</u>	33.4 <u>t</u>	31.0 t	32.6 <u>t</u>	32.2 t	32.2 t
C-9	42.8 <u>d</u>	41.0 <u>d</u>	45.5 <u>d</u>	44.2 d	36.9 <u>d</u>	43.5 <u>d</u>	C-24	69.2 <u>s</u>	69.5 <u>s</u>	150.3 <u>s</u>	157 3 <u>s</u>	68.3 <u>s</u>	68.0 <u>s</u>
C-10	50.4 <u>s</u>	52.2 <u>s</u>	53.9 <u>s</u>	128.0 <u>s</u>	47.2 <u>s</u>	47.4 <u>s</u>	C-25	138.9 <u>s</u>	139.1 <u>s</u>	122.8 5	122.1 s	137.9 <u>s</u>	137.8 8
C-11	23.6 <u>t</u>	22.6 <u>t</u>	22.7 <u>t</u>	26.8 <u>t</u>	21.6 <u>t</u>	22.5 <u>t</u>	C-26	165.2 <u>s</u>	165.4 <u>s</u>	166.8 <u>s</u>	164.9 <u>s</u>	164.2 <u>s</u>	164.2 8
C-12	39.6 <u>t</u>	39.9 <u>t</u>	39.1 <u>t</u>	38.7 <u>t</u>	38.4 <u>t</u>	38.5 <u>t</u>	C-27	129.7 <u>t</u>	130.0 <u>t</u>	12.3 g	58.1 <u>t</u>	128.8 t	128.8 t
C-13	42.5 <u>s</u>	43.1 <u>s</u>	42.5 <u>s</u>	43.0 <u>s</u>	41 5 5	41.5 <u>s</u>	C-28	25.6 g	25.7 g	20 6 g	20.7 g	24.5 g	24.5 g
C-14	56.0 <u>d</u>	56.0 <u>d</u>	56.3 <u>d</u>	55.1 d	55 1 <u>d</u>	54.6 ₫	-OAc				21.2 g		_
C-15	24.0 <u>t</u>	24.2 <u>t</u>	24.3 <u>t</u>	23 9 <u>t</u>	22.9 <u>t</u>	22 9 t					21.0 g 20.9 g		

* 25 MHz ** Carbon numbering is based on withametelin, i.e., carbons bearing -OAc and methyl are C-1 and C-10 respectively *** Not detected

Beside withametelins, the only other withanolides known to bear bridged bicyclic side chains are achistins¹⁷ and the only difference in the side chain of these two groups of compounds is that while in achistins C-21 and C-24 are directly linked, in withametelin they are separated by an oxygen bridge. Biogenetically, however, there exists a commonness in these two systems which are considered to be formed by S_N^2 ' type reaction of appropriately substituted withanolide (Scheme II). Daturametelin B ($\frac{6}{2b}$), isolated from the methanolic extract of fresh leaves of D. metel by Japanese workers 18 might very well be the precursor of withametelin but the suggestion that withametelin is an artefact formed during chromatography over silica gel column is not tenable because withametelin was isolated from the petroleum ether extract of the plant material in a fairly good yield (0.1%) and daturametelin B (6b), a glucoside, is not likely to be extracted with this solvent. The alternative suggestion¹⁹ that daturametelin B ($\underline{6b}$), a constituent of fresh leaves of <u>D</u>. <u>metel</u>, is transformed to withametelin during drying of the plant material is, no doubt, a possibility but our observation based on HPLC analysis, suggests that both daturametclin B ($\underline{6b}$) and withametelin coexist in fresh as well as in dry leaves, and that the former is the likely precursor of the latter.

EXPERIMENTAL :

M.ps were taken on a TOSHNIWAL melting point apparatus and are uncorrected. Column chromatography was carried out over silica gel (60-120 mesh, B.D.H.) and TLC over silica gel G (B.D.H.). All the samples were routinely dried over P_2O_5 at 80-140° (depending upon the m.ps of the compounds) for 24 hrs <u>in vacuo</u> and were tested for purity by TLC. Optical rotations were measured on a JASCO DIP-360 polarimeter and CD spectra on a JASCO A-3 instrument. IR

spectra were recorded on a SHIMADZU IR-27G spectrometer, UV spectra on a SHIMADZU UV-202 spectrophotometer, 1 H and 13 C NMR spectra on a JEOL FX-90Q, JEOL JNM FX-100 and JNM FX-500 spectrometers (TMS as internal standard). Low resolution EIMS were determined with a HITACHI M-52 and high resolution EIMS with a JEOL-01-SG-2 spectrophotometers.

Isolation of withametelin (1) and isowithametelin (2): Air dried and pulverized leaves of <u>D</u>. <u>metel</u> (600 g) were extracted with petroleum ether (80-100°) in a soxhlet apparatus for 20 hrs. The petroleum ether extract (25 g) was chromatographed over silica gel (125 g) and eluted initially with petroleum ether followed by C_6H_6 . C_6H_6 fractions (4 g) on rechromatography over EtOAc washed alumina (40 g) and elution with petroleum ether-EtOAc (17:3) afforded withametelin (600 mg). Isowithametelin was isolated by HPLC of the mother liquor of withametelin over silica gel column (No. S.I.G., I.D. 22 mm). Elution of the column with <u>n</u>-hexane-EtOAc (3:1) at the rate of 9 ml/min afforded isowithametelin after 20 min and withametelin after 22 min.

Withametelin ($\underline{1}$) crystallized from EtOAc as colourless needles, m.p. 210°; $[\alpha J_{D} - 64.4^{\circ} (\underline{c} 0.45, CHCl_{3})$; HREIMS $\underline{m}/\underline{z}$: 436.2426 (M⁺); EIMS $\underline{m}/\underline{z}$: 436 (M⁺), 421, 354, 268, 253, 225, 173, 147, 145, 131, 107, 105, 91, 67, 55; UV (MeOH) λ_{max} nm: 222 (\underline{c} 7000); IR (KBr) ν_{max} cm⁻¹: 1718, 1655; ¹H NMR (500 MHz, CDCl_{3}) & 6.77 (1H <u>ddd</u>, J=9.8, 5.1, 2.5 Hz, H-3), 6.76 (1H <u>br</u> <u>s</u>, H-27), 6.02 (1H <u>br</u> <u>s</u>, H-27), 5.88 (1H <u>dd</u>, J=9.8, 2.5 Hz, H-2), 5.58 (1H <u>t</u>, J=5.8 Hz, H-6), 4.65 (1H <u>br</u> <u>s</u>, W/2=8.8 Hz, H-22), 3.95 (1H <u>d</u>, J=12.7 Hz, H-21), 3.73 (1H <u>dd</u>, J=12.7, 3.0 Hz, H-21), 3.30 (1H <u>dt</u>, J=20.0, 2.5 Hz, H-4\beta), 2.84 (1H <u>dd</u>, J=20.0, 5.1 Hz, H-4\alpha), 1.44, 1.22, 0.71 (3H <u>s</u> each, H₃-28, -19, -18); Found: C, 76.42; H, 8.09, calc. for C₂₈H₃₆O₄: C, 77.03; H, 8.31%.

Isowithametelin ($\underline{2}$), m.p. 280°; $\lceil \alpha \rceil_{D} -77.7°$ (\underline{c} 0.26, CHCl₃); EIMS $\underline{m}/\underline{z}$: 436 (M⁺); UV (MeOH) λ_{max} nm: 233, 227sh (ε 13000); IR (CHCl₃) ν_{max} cm⁻¹: 1720, 1714; ¹H NMR (500 MHz, CDCl₃) δ : 6.73 (1H \underline{s} , H-27), 6.02 (1H \underline{dd} , J=10.0, 2.5 Hz, H-4), 5.99 (1H \underline{s} , H-27), 5.62 (1H \underline{dd} , J=5.0, 2.5 Hz, H-6), 5.58 (1H \underline{dt} , J=10.0, 5.0 Hz, H-3), 4.63 (1H \underline{br} \underline{s} , W/2=10.0 Hz, H-22), 3.89 (1H \underline{d} , J=13.0 Hz, H-21), 3.70 (1H \underline{dd} , J=13.0, 2.0 Hz, H-21), 3.28 (1H \underline{d} , J=20.0, 5.0 Hz, H-2 α), 1.42, 1.33, 0.68 (3H \underline{s} each, H₃-28, -19, -18).

<u>Hydrogenation of withametelin to 2,3,5,6,25,27-hexahydrowithametelin</u> <u>and 5</u>: Withametelin (100 mg) was hydrogenated in methanolic solution using 5% Pd-C (30 mg) as a catalyst for 4 hrs. Usual work-up followed by purification over silica gel column yielded hexahydrowithametelin (55 mg) and 5 (10 mg). Hexahydrowithametelin, m.p. 258°; $[\alpha J_{p}]$ -82.7° (<u>c</u> 0.26, CHCl₃); EIMS <u>m/z</u>: 442 (M⁺), 424, 369, 351, 319, 271, 231, 147, 124; IR (KBr) $\lambda_{\max} cm^{-1}$: 1725, 1690; ¹H NMR (100 MHz, CDCl₃) & 4.58 (1H <u>br</u> <u>s</u>, H-22), 3.86, 3.46 (1H <u>br</u> <u>d</u>, J=13.0 Hz each, H₂-21), 1.32 (3H <u>d</u>, J=7.0 Hz, H₃-27), 1.20, 1.12, 0.64 (3H <u>s</u> each, H₃-28, -19, -18); <u>5</u>, m.p. 278°; /α/_D -68.4° (<u>c</u> 0.36, CHCl₃); EIMS <u>m/z</u>: 440 (M⁺), 355, 152, 150, 125; UV (MeOH) λ_{\max} nm: 228 (ε 9200); ¹H NMR (500 MHz, CDCl₃) & 5.42 (1H <u>d</u>, J=6.0 Hz, H=6), 4.37 (1H <u>dt</u>, J=13.5, 3.6 Hz, H=22), 4.00 (1H <u>br</u> <u>d</u>, J=11.8 Hz, H=21), 3.74 (1H <u>dd</u>, J=11.8, 5.0 Hz, H=21), 1.93, 1.88, 1.24, 0.70 (3H <u>s</u> each, H₃-28, -27, -19, -18); CD (MeOH): [θ]₂₅₀ +10700.

 $\begin{array}{c} \mbox{Transformation of withametelin to secowithametelin (<u>6</u>): A solution of withametelin (100 mg) in MeOH (15 ml) was treated with a few drops of HClo₄ and left overnight at room temp. The reaction product on chromatography over silica gel furnished white plates of secowithametelin (60 mg) from petroleum ether-EtOAc (3:1) eluates, m.p. 190°; / α /_D +28.5° (<u>6</u> 0.75, CHCl₃); UV (MeOH) λ_{max} nm: 228 (£ 14040); IR (KBr) ν_{max} cm⁻¹: 3490, 1718, 1684; ¹H NMR (100 MHz, CDCl₃) δ: 6.73 (1H <u>ddd</u>, J=10.0, 5.5, 2.5 Hz, H=3), 5.85 (1H <u>dd</u>, J=10.0, 2.5 Hz, H=2), 5.53 (1H <u>d</u>, J=6.0 Hz, H=6), 4.49 (1H <u>dt</u>, J=12.0, 4.0 Hz, H=2), 4.24 (2H AB <u>g</u>, J=12.0 Hz, <math>\Delta v$ 16.0 Hz, H₂-27), 3.94 (2H <u>m</u>, H₂-21), 3.38 (3H <u>s</u>, -OMe), 2.08, 1.23, 0.75 (3H <u>s</u> each, H₃-28, -19, -18); CD (Dioxane): [θ]₂₅₃ +13540, [θ]₃₂₀ -8000.

 $\frac{\text{Conversion of } 3 \text{ to } 8: \text{ Compound } 3 \text{ (20 mg) in AcOH (1.5 ml) was treated with 30%}}{\text{HClO}_4 (0.2 ml) at room temp. and left overnight. Usual work-up followed by chromatography over silica gel yielded 8 as yellow needles, UV (MeOH) <math>\lambda_{\max}$ nm: 312, 219 (£ 2509, 14200).

 $\begin{array}{c} \underline{\text{Epoxidation of withametelin to 9 and 10}: A solution of withametelin (50 mg)} \\ \text{in CHCl}_{3} (5 \text{ ml}) \text{ was treated with } \underline{\text{m-chloroperbenzoic acid (50 mg) at room temp. for 4 hrs.} \\ \text{Usual work-up yielded a mixture of epoxides which was resolved by HPLC to give 9 (20 mg) as a major product, m.p. 218° (dec); EIMS <math>\underline{\text{m/z}}: 452 \text{ (M}^+); {}^{1}\text{H} \text{ NMR (90 MHz, CDCl}_{3}) \& 6.75 \text{ (1H } \underline{\text{s}}, \text{H-27}) \\ 6.62 \text{ (1H } \underline{\text{m}}, \text{H-3}), 6.01 \text{ (1H } \underline{\text{s}}, \text{H-27}), 5.85 \text{ (1H } \underline{\text{m}}, \text{H-2}), 4.64 \text{ (1H } \underline{\text{br}} \underline{\text{s}}, \text{H-22}), 3.80 \text{ (2H AB } \underline{\text{g}}, \\ J=20.0 \text{ Hz}, \text{ H}_{2}-21), 3.02 \text{ (1H } \underline{\text{t}}, J=6.0 \text{ Hz}, \text{H-6}), 1.44, 1.33, 0.66 \text{ (3H } \underline{\text{s}} \text{ each}, \text{ H}_{3}-28, -19, -18); \\ \text{CD (Dioxane): } \theta_{345} - 4565; \underline{10} \text{ (7 mg) as a minor product, m.p. 228° (dec); EIMS } \underline{\text{m/z}} \cdot 452 \text{ (M}^+); \\ {}^{1}\text{H} \text{ NMR (90 MHz, CDCl}_{3}) \& 6.6.86 \text{ (1H } \underline{\text{dd}}, \text{J}=10.0, 5.0, 2.5 \text{ Hz}, \text{H-3}), 6.74 \text{ (1H } \underline{\text{s}}, \text{H-27}), 6.02 \\ \text{(1H } \underline{\text{dd}}, \text{J}=10.0, 2.5 \text{ Hz}, \text{H-2}), 6.00 \text{ (1H } \underline{\text{s}}, \text{H-27}), 4.63 \text{ (1H } \underline{\text{br}} \underline{\text{s}}, \text{H-22}), 3.10 \text{ (1H } \underline{\text{br}} \underline{\text{s}}, \text{H-6}), \\ \textbf{1.44, 1.24, 0.67 (3H } \underline{\text{s}} \text{ each}, \text{ H}_{3}-28, -19, -18); CD \text{ (Dioxane): } [\theta]_{345} + 2983. \\ \end{array}$

<u>Selenium dioxide oxidation of withametelin to 3, 3a, 11 and 12</u>: A solution of withametelin (250 mg) and SeO₂ (90 mg) in 3 ml $C_{6}H_{6}$ and 6 ml AcOH was refluxed for 8

hrs. Usual work-up followed by silica gel column chromatography of the product afforded <u>3</u> (45 mg), <u>3a</u> (20 mg), <u>11</u> (20 mg) and <u>12</u> (5 mg). <u>3a</u>, m.p. 278°; ¹H NMR (80 MHz, CDCl₃) & 6.92 (1H <u>dd</u>, J=9.6, 6.1 Hz, H-3), 6.75 (1H <u>s</u>, H-27), 6.14 (1H <u>d</u>, J=6:1 Hz, H-4), 6.03 (1H <u>d</u>, J=9.6 Hz, H-2), 6.00 (1H <u>s</u>, H-27), 4.62 (1H <u>br</u> <u>s</u>, H-22), 4.59 (1H <u>br</u> <u>s</u>, H-6), 3.80 (2H AB <u>g</u>, J=14.0 Hz, H₂-21), 1.47, 1.41, 0.76 (3H <u>s</u> each, H₃-19, -28, -18); <u>11</u>, m.p. 328°(dec); UV (MeOH) λ_{max} nm: 207 (ε 9744); ¹H NMR (90 MHz, CDCl₃) & 6.75 (1H <u>s</u>, H-27), 6.74 (1H <u>d</u>, J=10.3 Hz, H-3), 6.46 (1H <u>d</u>, J=10.3 Hz, H-2), 6.00 (1H <u>s</u>, H-27), 4.62 (1H <u>br</u> <u>s</u>, H-22), 3.70 (2H AB <u>g</u> with fine splitting, J=12.6 Hz, H₂-21), 1.44, 1.40, 0.68 (3H <u>s</u> each, H₃-28, -19, -18); <u>12</u>, UV (MeOH) λ_{max} nm: 355, 207 (ε 3986, 19395); ¹H NMR (90 MHz, CDCl₃) & 6.96 (1H <u>dd</u>, J=10.0, 6.0 Hz, H-3), 6.76 (1H <u>s</u>, H-27), 6.16 (1H <u>d</u>, J=10.0, 2.5 Hz, H-4), 6.00 (1H <u>s</u>, H-27), 6.00-5.80 (3H <u>m</u>, H-2, -6, -7), 4.63 (1H <u>br</u> <u>s</u>, H-22), 3.80 (2H AB <u>q</u> with fine splitting, J=12.6 Hz, H₂-21), 3.80 (2H AB <u>q</u> with fine splitting, J=12.6 Hz, H₂-21), 1.44, 1.24, 0.78 (3H <u>s</u> each, H₃-28, -19, -18).

Diisobutylaluminium hydride reduction of withametelin to <u>13</u>: A solution of withametelin (27 mg) in dry tetrahydrofuran (1 ml) was treated with 0.2 ml of DIBAH (1M/1) at -30° for 1 hr. Acidic work-up (NH₄Cl+3N HCl) of the reaction mixture and purification by silica gel column yielded <u>13</u> (15 mg), EIMS $\underline{m/z}$: 424 (M⁺), 406, 109; IR (CHCl₃) v_{max} cm⁻¹: 3450, 1460, 1395, 1100, 1000, ¹H NMR (90 MHz, CDCl₃) δ : 5.72-5.20 (3H m, H-2, -3, -6), 5.00 (1H <u>s</u>, H-26), 4.02-3.82 (2H m, H-1, -22), 3.50 (2H <u>t</u>, J=12.0 Hz, H₂-21), 1.75, 1.64, 0.99, 0.76 (3H <u>s</u> each, H₃-27, -28, -19, -18).

 $\frac{\text{Conversion of isowithametelin to 6a}{(10 \text{ mg}) \text{ in MeOH (1.5 ml) with 1 drop of 30% HClo}_4 \text{ and a work-up same as that applied in case of secowithametelin (<u>6</u>), yielded a compound <u>6a</u> (4 mg), EIMS <u>m/z</u>: 468 (M⁺); UV (MeOH) <math>\lambda_{\text{max}}$ nm: 227 (ε 16300); ¹H NMR (90 MHz, CDCl₃) δ : 5.97 (1H <u>br</u> <u>d</u>, J=9.2 Hz, H-4), 5.66 (1H <u>br</u> <u>d</u>, J=6.0 Hz, H-6), 5.56 (1H <u>dd</u>, J=9.2, 4.0 Hz, H-3), 4.41 (1H <u>dt</u>, J=12.6, 3.9 Hz, H-22), 4.16 (2H AB <u>q</u>, J=11.0 Hz, H₂-27), 3.94 (2H <u>m</u>, H₂-21), 3.31 (3H <u>s</u>, -OMe), 2.00, 1.29, 0.69 (3H <u>s</u> each, H₃-28, -19, -18); CD (Dioxane) [θ]₂₄₁ +32600.

 $\frac{\text{Transformation of withametelin to isowithametelin:} A solution of withametelin (25 mg), ethylene glycol (0.2 ml) and p-toluenesulphonic acid (2 mg) in C₆H₆ (5 ml) was heated at 110° for 24 hrs. Usual work-up and purification gave <u>15</u> (10 mg), EIMS <u>m/z</u> 480 (M⁺); ¹H NMR (90 MHz, CDCl₃) & 6.68 (1H <u>s</u>, H-z/), 6.02 (1H <u>s</u>, H-27), 5.97 (1H <u>dd</u>, J=9.0, 2.0 Hz, H-4), 5.66 (1H <u>br</u> <u>d</u>, J=6.0 Hz, H-6), 5.48 (1H <u>m</u>, H-3), 4.66 (1H <u>br</u> <u>s</u>, H-22), 4.00 (4H <u>br</u> <u>s</u>, -0-CH₂-CH₂-O-), 3.84 (2H AB <u>q</u>, J=12.6 Hz, H₂-21), 2.28 (2H <u>m</u>, H₂-2), 1.44, 1.14, 0.72 (3H <u>s</u> each, H₃-28, -19, -18). Compound <u>15</u> (8 mg) was treated with <u>p</u>-toluenesulphonic acid (2 mg) in aq. acetone for 12 hrs at room temp. Work-up and purification gave <u>2</u> (5 mg) indistingui-$

shable from isowithametelin in all respects (t.l.c., m.m.p., mass, IR, ¹H NMR).

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REFERENCES :

- 1. Part 19 of withanolides.
- Oshima, Y.; Bagchi, A.; Hikino, H.; Sinha, S. C.; Sahai, M.; Ray, A. B. <u>Tetrahedron</u> Lett. 1987, 28, 2025-2028.
- Frolow, F.; Ray, A. B.; Sahai, M.; Glotter, E.; Gottlieb, H. E.; Kirson, I. J. Chem. Soc. Perkin Trans. I 1981, 1029-1032.
- 4. Velde, V. V., Lavie, D. Phytochemistry 1982, 21, 731-733.
- a) Sinha S. C. <u>Isolation and Structure Determination of C₂₈-steroids of Solanaceous</u> <u>Plants</u>, Ph.D. thesis, Banaras Hindu University, 1987. b) Sinha, S. C.; Ray, A. B. <u>J. Ind. Chem. Soc. 1988</u>, <u>65</u>, 740.
- a) Alkyoni, I. <u>Chem. Ber</u>. 1962, <u>95</u>, 279-280. b) Robinson, D. J.; Sosnovsky, S. Synthesis 1973, 567-603.
- 7. Sahai, M.; Kırson, I. J. Nat. Prod. 1984, 47, 527-529.
- 8. Kundu, S.; Sinha, S. C.; Bagchi, A; Ray, A. B. Phytochemistry 1989, 28, in press.
- 9. Gottlieb, H. E.; Kirson, I. Org. Magn. Reson. 1981, 16, 20-25.
- Ray, A. B.; Alı, A.; Sahaı, M.; Schiff, P. L., Jr.; Knapp, J. E.; Slatkin, D. J. Chemistry & Industry (London) 1981, 62-64.
- 11. Djerassi, C. Steroids Reactions 1963, p. 371, Holden Day Inc., San Francisco.
- 12. Stothers, J. B. Carbon-13 NMR Spectroscopy 1972, p. 447, Academic Press, New York.
- Glotter, E.; Kırson, I.; Lavie, D.; Abraham, A. <u>The Withanolides, A Group of Natural</u> <u>Steroids in Bio-organic Chemistry</u> 1978, Vol II, pp. 57-95, ed. E. E. van Tamelen, Academic Press, New York.
- 14. Glotter, E.; Kırson, I.; Abraham, A.; Lavie, D. Tetrahedron 1973, 29, 1353-1364.
- Siddiqui, S.; Sultana, N.; Ahmad, S. S., Haider, S. I. <u>Phytochemistry</u> 1987, <u>26</u>, 2641-2643.
- 16. Velde V. V.; Lavie, D. Phytochemistry 1981, 20, 1359-1364.
- Usubillaga, A.; Castellano, G. de.; Zabel, V.; Watson, W. H. J. Chem. Soc. Chem. Comm. 1980, 854-855.
- Singu, K.; Kajimoto, T.; Furusawa, Y.; Nohara, T. <u>Chem. Pharm. Bull</u>. 1987, <u>35</u>, 4359-4361.
- 19. Personal communication from Prof. T. Nohara of Kumamoto University, Japan.